Field of the invention

peptide, Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with cyclodextrin derivative. More particularly, the invention relates to Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with cyclodextrin derivatives such as beta-cyclodextrin, hydroxypropyl-beta cyclodextrin, hydroxyethyl-beta-cyclodextrin. Dimethyl-beta cyclodextrin. The invention also relates to a process for the preparation of pharmaceutical compositions containing the inclusion complexes of opiod peptide and the use thereof in the treatment of alleviating pain and acute inflammation, which can be used as a substitute for paraestic-analaesies.

Background of the invention

be utilized as substitute for the narcotic analgesiscs, currently being used, having improved biopharmaceutical properties such as low toxicity, lesser tolerance, longer duration of action and least abuse potential. The development of the peptide as drug is restricted due to its poor oral efficacy. In view of this, it is essential to develop orally active formulations. There has been tremendous emphasis on the development of innovative strategies for the oral delivery of peptide based drugs, with increased water solubility, dissolution, bioavailability and improved oral efficacy.

Prior Art

Soon after the discovery of enkephalin by Huges et. Al [Huges et. Al, Nature 258, 577 (1975)]. an endogenously occurring pentapeptide with morphinomimetic activity, structure activity relationship studies were undertaken world-wide with the objective of getting a synthetic congener that would be clinically acceptable pain killer as substitute to morphine. Since the analgesia evoked by enkephalins was only weak and transient following their administration by intra-cerebral route, greater emphasis was laid on the structural modification of the penta peptide which would lead to the peptide(s) capable of eliciting profound analgesis even after their systemic administration.

studies so far, are the Sandoz compound FK –33-824 [Tyr-D-Ala-Gly-Met(o)-0] and the Lilly compound met-keohamid (Tyr-D-Ala-Gly-Phe-MeMet-NH2. [Von Graffenreid, B., del Pozo, E., Roubicek, J., Krebs, E., Poldinger W., Burmeister, P. and Kerp, L., Nature, 272, 729 (1978) and Frederickson, R.C.A., Smithwick, E. L., Shuman, R. and Bemis, K.G., Science, 211, 603, (1981) Frederickson, R.C.A.. In 'Opioid Peptides: Molecular Pharmacology, Bio synthesis and analysis' Rapaka, R.S. and Hawks, R. L. eds. NIDA Research Monograph, 70 367. (1986) FK-33-824 gave only slight preference for μ-receptors and met-kephamid was found essentially pon-elective for μ and δ receptors. A strong analgesic effect was exerted by both the compounds by systemic route of administration. However, due to a number of serious side effects produced by FK –33-824 therefore it was no longer pursued for further developments as candidate analgesic drug Relatively fewer side effects were observed with met-kephamid, but due to the hypotensive effect this compound was also finally abandoned

Another enkephalin analogs Tyr-D-Mei-Gly-Phe-Pro-NH2 (Flodes, J., toros II., Borvenderg, J., Karezag, I., Tolna, J., Marosfi, S., Varadi, A., Gara, A., Ronai, A.Z. and Szilaggi, G. Life Sciences 33, Supp. 1, 769 (1983)] and Tyr-Arg-Gly-Phe(p-NCy)-Pro-NH2 (BW-443C) [Follenfant, R.L., Hardy, G.W., Lowe, L.A., Schneider, C. and Spitth, T.W. Br. J. Pharmacol. 93, 85, (1988) and Kriss, M.G., Gralla, R.J., Clark, R.A., Tyson, L.B. and Groshen, S., J. Clin. Onclol., 6, 663, (1988)] were also shown to be more potent analgesic than morphine but due to number of side effects these compounds were also dropped after initial clinical trials. Similarly, structure activity relationship studies were undertaken in our laboratory and an enkephalin analog Tyr-D-Ala-Gly-MePhe-Gly-NHC3H7 [Raghubir, R., Patnaik, G.K., Sharma, S.D., Mathur, K.B. and Dhawan B.N., In recent progress in chemistry and biology of centrally acting peptides. Dhawar B.N. and Rapaka R.S. eds., 167, (1988) and Indian Patent no. 173568 19.10.1989] synthesised earlier in our laboratory and found to be more potent than morphine following systemic administration. This is a highly μ-receptor selective in central and peripheral assay and produces highly profound and long lasting analgesia. (C. Nath, G.K. Patnaik, W. Haq, K.B. Mathur, R.C. Srimal, B.N. Dhawan and F. Porreca. Pharmacological Research., 31, 269-273, 1995) The compound and its process for the synthesis was first disclosed in Indian patent [Indian Patent no. 173568 19.10.1989]. Subsequently chronic and subacute toxicity studies on this compound were carried out and it is now disclosed that this compound is safe and did not produce any noticeable toxic side effects. This compound was also studied for their addiction liabilities and tolerance and found to elicit significantly reduced tolerance and addiction properties as compared to morphine. The compound is virtually devoid of any major CNS effects like sedation and respiratory depression; it is also virtually devoid of any significant cardiovascular effects. Therefore, this compound has a potential as centrally active analgesic agent, which can be used as a substitute to/narcotic analgesics (Morphine and related substances). However, this compound upon oral/administration produced poor response and extremely high dose is required to obtain symilar magnitude of response as observed after parenteral administration owing to its decomposition and poor absorption. The oral efficacy of the therapeutic agents is considered to be highly desirable, therefore, inspite of a profound analgesia and favourable pharmacological effect and almost devoid of toxic effects, the development of the peptide as drug is restricted due to its poor oral efficacy. In view of this, it is essential to develop orally active formulations.

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there has been tremendous emphasis, on the development of innovative for the oral delivery of peptide based drugs, (A Fasano. (1998) strategies 16, 152-157) with increased water solubility dissolution, TIBTECH., bioavailability and improved oral efficacy (Z. Shao, 1992). Cyclodextrins are reported in the literature that they increase water solubility, dissolution, bioavailability and stability of compound by forming inclusion complexes. [R. Krishnamoorthy and A. K. Mitra, Pharma. Res., 9: 1157-1163 (1992)]. Recently it was reported in the literature that the β-cyclodextrin inclusion complex increase the half life of Leu-enkephalin from 45 min to 75 min in case of enzyme hydrolysis with leucine amino peptidase (W. J. Erwin., A. K. Dwivedi, P.A. Holbrook, and M. J. Dey, Pharma Res., 11, 1994, 1698-1703). Therefore, the preparation of inclusion complex of peptides and other substances with cyclodextrin are reported in the literature. The advantage of binding substances into inclusion complexes with cyclodextrin is also known in other substances. US Patent No.4603123, 5840714 and 5855916 disclosed the increased therapeutic efficacy and reduced toxic effects of piroxicam, ibuproxam and acid base type drugs respectively.

Objects of the invention

The main object of the present invention is to provide novel L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide: cyclodextrin complexes having significantly improved oral efficacy and prolonged duration of action.

Another object is to provide novel inclusion complexes that can be utilised as substitute for narcotic analgesics, said complexes having improved biopharmaceutical properties such as low toxicity, lesser tolerance, longer duration of action and least abuse potential.

Still another object is to provide novel L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide complexes with increased water solubility, dissolution, bioavailability and improved oral/transdermal efficacy.

Yet another object is to provide pharmaceutical compositions containing said novel inclusion complexes having improved analgesic activity with longer duration of activity and improved efficacy.

Still another object is to provide methods for the treatment of acute inflammatory conditions employing the said inclusion complexes.

Summary of the invention

The above and other objects are realized by the present invention that provides novel inclusion complexes having improved analgesic property and longer duration of activity. The invention also provides pharmaceutical compositions containing said inclusion complexes and methods of treatment employing such inclusion complexes.

Detailed Description:

Accordingly, the invention provides novel inclusion complexes having significantly improved oral efficacy and prolonged duration of action selected from the group consisting of highly potent opioid peptide of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with a cyclodextrin derivative wherein said derivative is selected from the group consisting of beta cyclodextrin, hydroxypropyl-beta cyclodextrin, dimethyl-beta cyclodextrin and hydroxyethyl-beta-cyclodextrin

In an embodiment, the invention provides cyclodextrin derivative selected from beta cyclodextrin, hydroxypropyl-beta cyclodextrin, dimethyl-beta cyclodextrin, hydroxyethyl-beta-cyclodextrin.

In yet another embodiment, the inclusion complex comprises L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta,-cyclodextrin.

He still another embodiment, the inclusion complex comprises L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with hydroxyehtyl beta-cyclodextrin-

In yet another embodiment, the inclusion complex comprises L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with hydroxypropyl beta-cyclodextrin.

In another embodiment, the inclusion complex comprises L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with dimethyl beta, cyclodextrin.

In still another embodiment, the molar ratio between L-Tyrosyl-D-alanyl-glycy-N-methylphenylalanyl-glycyl-isopropylamide and said cyclodextrin derivative is 1:5 to 2:1.

In yet another embodiment, the invention provides pharmaceutical compositions comprising a therapeutically effective amount of inclusion complex of L-Tyrosyl-D-alanyl-glycy-N-methylphenylalanyl-glycyl-isopropylamide with beta-cyclodextrin having improved analgesic activity with longer duration of action as compared with the free peptide.

In yet another embodiment, the pharmaceutical composition has potential for clinical application as an analgesic.

In still another embodiment, the pharmaceutical composition is formulated in various physical forms such as tablets, injections, capsules.

The invention also provides methods for the treatment of acute inflammations and for alleviating pain comprising the step of administration of pharmaceutical composition containing the inclusion complex to a patient in need thereof.

In an embodiment, the inclusion complex is administered orally or transdermally or rectal route.

In yet another embodiment, the pharmaceutical composition exhibits significant analgesic activity with reduced dependence liability, respiratory depression, gastric irritation and sedation.

In another embodiment, the method for the treatment of acute inflammations and for alleviating pain, comprises oral administration of inclusion complex of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta-cyclodextrin.

In another embodiment, the method for the treatment of acute inflammations and for alleviating pain, comprises topical application of inclusion complex of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta cyclodextrin.

The L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide: cyclodextrin (1.2) complex of the invention is used for the oral delivery of the peptide as these complexes provide more protection to the peptide against the gastric environment. The L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide: cyclodextrin (1:1) complex is used for the transdermal delivery of the peptide as these complexes facilitate the transdermal permeation of the peptide.

TECHNICAL SOLUTION

The invention is illustrated by the following Examples which in no way represent a limitation thereof

EXAMPLE 1

Preparation of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide [Figure 1]:

The penta-peptide was synthesised by solution phase method of peptide synthesis employing three plus two fragment condensation method or by step up chain elongation procedure. This compound was also prepared on solid support employing commercially available supports using either BOC or FMOC chemistry. The coupling reactions were performed by commonly available reagents adopting reported procedures. The out line of the process for the preparation of the penta-peptide has been disclosed in Indian Patent No. 173568 19.10.1989.

FIG. 2 shows Mass spectrum of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide.

FIGS. 3 show NMR spectrum of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide.

FIG. 4 shows DSC (differential scanning calorimetry) thermogram of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide.

EXAMPLE 2

Preparation of inclusion complex of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta-cyclodextrin

a) Procedure in aqueous medium

Beta-cyclodextrin (1.135 g, 1.0 m mole) in water (25 ml) was heated to boiling temperature. L-Tyrosyl-D-alanyl-glycyl-N-methylphenyl-alanyl-glycyl-isopropylamide (0.568 g, 1.0 m mole) was added into this solution, vigorously stirred for 15 minutes and the mixture was cooled during stirring to a temperature between 0 degree and 5 degree centigrade. The obtained complex was dried in vacuum at the temperature of about 40 degree C. Inclusion complex (1.62 g, 95.1%) of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta-cyclodextrin was obtained in the form of a white powder in the molar ratio of 1:1.

Path on reaction yields, L-Tyrosyl-D-alanyl-glycyl-N-methylphenyl-alanyl-glycylIsopropylamide content in the emplex (determined theoretically and experimentallyspectrophotometric determination at the wavelength of 275 nm) are summarised in Table 1Fig. 5 shows Differential scanning calorimetry (DSC thermogram) in the curve of the obtained product. There was not detected any endothermic transition for a melting point, characteristic of a physical mixture of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide.

Fig. 6 shows NMR spectrum of the complex of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta-cyclodextrin.

FIG. 7 shows Mass spectrum of the complex of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta-cyclodextrin.

b) Procedure in a solvent mixture (methanol/water in the ratio 5:20)

Beta.-cyclodextrin (1.135 g; 1.0 m mole) was dissolved in water (25 ml) at a temperature of about 70 degree C. During stirring a solution of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide (0.568 g; 1.0 m mole) in methanol (5 ml) was added. It was stirred for another 5 minutes, the solvents were evaporated and the obtained complex was dried in vacuum at a temperature about 40 degree C. Inclusion complex (1.59 g, 93.4%) of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta-cyclodextrin was obtained in the form of a white powder in the molar ratio of 1:1.

Differential scanning calorimetry, mass and NMR spectrum showed the same results as in the process for preparing the inclusion complex in an aqueous medium.

EXAMPLE 3

Preparation of inclusion complex of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta-cyclodextrin

a) Procedure in aqueous medium

Beta-cyclodextrin (1.135 g; 1.0 m mole) in water (25 ml) was heated to boiling temperature, into the boiling solution of L-Tyrosyl-D-alanyl-glycyl-N-methylphenyl-alanyl-glycyl-isopropylamide (0.284 g; 0.5 m mole) was added, vigorously stirred for 15 minutes and the mixture was cooled during stirring to a temperature between 0 degree and 5 degree C. The obtained complex was dried in vacuum at the temperature of about 40 degree C. Inclusion complex (1.33 g; 93.7%) of L-Tyrosyl-D-alanyl-glycyl-N-methylphenyl-alanyl-glycyl-isopropylamide with beta-cyclodextrin was obtained in the form of a white powder in the molar ratio of 1:2.

Data on reaction yields, L-Tyrosyl-D-alanyl-glycyl-N-methylphenyl-alanyl-glycyl-isopropylamide content in the complex (determined theoretically and experimentally-spectrophotometric determination at the wavelength of 275 nm) are summarised in Table 1. Fig. 8 shows Differential scanning calorimetry (DSC thermogram) in the curve of the obtained product there was not detected any endothermic transition for a melting point, characteristic of a physical mixture of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide/beta-cyclodextrin.

Fig. 9 shows NMR spectrum of the complex of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta-cyclodextrin.

b) Procedure in a solvent mixture (methanol/water in the ratio 5:20)

Beta -cyclodextrin (1.135 g, 1.0 m mole) was dissolved in water (25 ml) at a temperature of about 70 degree C. L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide (0.284 g; 0.5 m mole) in methanol (5 ml) was added and stirred for another 5 minutes. The solvents were evaporated and the complex obtained was dried in vacuum at a temperature about 40 degree C. Inclusion complex (1.36 g, 95.8%) of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta -cyclodextrin was obtained in the form of a white powder in the molar ratio of 1:2.

Differential scanning calorimetry and NMR spectrum showed the same results as in the process for preparing the inclusion complex in an aqueous medium.

EXAMPLE 4

Preparation of inclusion complex of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with hydroxypropyl-beta-cyclodextrin

a) Procedure in methanolic medium

L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide (0.568 g; 1.0 m mole) was added to a solution of hydroxypropyl-beta-cyclodextrin (1.38 g; 1.0 m mole) in methanol (25 ml) and the obtained solution was stirred for another 5 minutes at room temperature. Methanol was then evaporated and the obtained complex was dried in vacuum at the temperature of 40 degree C. Inclusion complex (1.82g; 93.4%) of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with hydroxypropyl-beta-cyclodextrin was obtained in the form of a white powder in the molar ratio of 1:1.

Dexa on reaction yields, L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl glycyl-10 isopropylamide content in the complex (determined theoretically and experimentally

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spectrophotometric determination at the wavelength of 275 nm) of the consummarised in Table 1.

Fig. 10 shows differential scanning calorimetry (DSC thermogram)

Fig 11 shows NMR spectrum of the complex of L-Tyrosyl-D-alanyl-glycyl-Nmethylphenylalanyl-glycyl-isopropylamide with hydroxypropyl-beta-cyclodextrin.

b) Procedure in aqueous medium

Hydroxypropyl-beta-cyclodextrin (1.38 g; 1.0 m mole) was dissolved in water (40 ml) and the obtained solution was heated to the temperature of 70 degree C and L-Tyrosyl-D-alanylglycyl-N-methylphenylalanyl-glycyl-isopropylamide (0.568 g; 1.0 m mole) was added. It was vigorously stirred for another 15 minutes and then the solution was filtered. The filtrate was frozen in liquid nitrogen and lyophilised. Inclusion complex (1.81g; 92.9 %) of L-Tyrosyl-Dalanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with hydroxypropyl-betacyclodextrin was obtained in the form of a white powder in the molar ratio of 1:1. Differential scanning calorimetry and NMR spectrum showed the same results as in the process for preparing inclusion complex in the methanolic medium.

EXAMPLE 5

Preparation of inclusion complex of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanylglycyl-isopropylamide with hydroxyethyl-beta-cyclodextrin

a) Procedure in methanolic medium

L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide (0.568 g; 1.0 mmole) was added to a solution of hydroxyethyl-beta-cyclodextrin (1.44 g; 1.0 mmole) in methanol (10 ml) and the obtained solution was stirred for another 5 minutes at room temperature. Methanol was then evaporated and the obtained complex was dried in vacuo at the temperature of 40 degree C. Inclusion complex (1.87 g; 93.1 %) of L-Tyrosyl-D-alanylglycyl-N-methylphenylalanyl-glycyl-isopropylamide with hydroxyethyl-.beta.-cyclodextrin was obtained in the form of a white powder in the molar ratio of 1:1.

Differential scanning calorimetry and NMR spectrum showed the formation of inclusion complex.

b) Procedure in aqueous medium

Hydroxyethyl beta eyclodextrin (1.44 g, 1.0 mmole) was dissolved in water (40 ml). The oblained solution was heated to the temperature of 70 degree. C. and L-Tyrosyl-D-alanylglycyl-N-methylphenylalanyl-glycyl-isopropylamide (0.568 g, 1.0 mmole) was added. It was vigorously stirred for another 15 minutes and then the solution was filtered. The filtrate was

D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with hydroxyethyl- beta-cyclodextrin was obtained in the form of a white powder in the molar ratio of 1:1.

Differential scanning calorimetry and NMR spectrum showed the formation of inclusion complex.

TABLE I

Reaction yield, L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide content in the complex for inclusion complex of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with different derivatives of beta-cyclodextrin.

Inclusion complex	Reaction yield (%)	% Content of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide in the complex
Beta-cyclodextrin (1:1)	95.1	97.2
Beta-cyclodextrin (1:2)	93.7	92.88
hydroxypropyl-beta-cyclodextrin (1:1)	93.4	98.7
hydroxyethyl-beta-cyclodextrin	93.1	97.5
(1:1)		

EXAMPLE 6

30 days toxicity study of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide in rat and rhesus monkey by subcutaneous route.

Compound L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide dissolved in saline was given daily by subcutaneous route to groups of rats (n=10 males and 10 females in each group) and rhesus monkeys (n=2 males and 2 females in each group) once daily for 30 consecutive days. The dose levels used in rats were 0.0, 7.5, 15.0 and 30.0 mg/kg and in monkeys 0.0, 3.75, 7.5 and 15 mg/kg. Dose levels were selected on the basis of rat ED of 3.0 mg/kg and extrapolation thereof in the monkey.

A careful general inspection of the animals was done at least once each day. Cageside observations included changes in skin and fur, eyes and mucous membranes, general activity and behavior and also general signs of abnormal functioning of respiratory, circulatory, autonomic and central nervous system and somatomotor activity.

Body weights (weekly in rats, initial and day 30 in monkeys), daily food (rat and monkeys) and water (rat only) consumption and parameters of hematology, urinalysis and blood biochemistry of the animals was recorded. All the animals were sacrificed at the end of 30 days. Necropsy was performed and histophathological examination of all the important organs and tissues were done.

Urinalysis included the recording of colour, specific gravity reaction (pH) and testing for sugar, acetone, urobillinogen, bilirubin, and albumin and occult blood. Microscopic examination of urinary sediment was done for the presence of epithelial cells, WBCs, RBCs, casts, crystals and other abnormal constituents.

The hematological examinations included hemoglobin, T-RBC count mean corpuscular hemoglobin concentration, mean corpuscular volume, total leukocyte count, packed cells volume mean corpuscular volume, platelet counts, differential leukocyte counts, erythrocyte sedimentation rate (monkeys only) and prothrombin time test (monkeys only).

Biochemical estimations in blood done at day 30 in rat, and day 0 and 30 in monkeys) included glucose, creatinine, urea nitrogen, sodium, potassium SGPT (ALT), alkaline phosphatase (ALP), bilirubin, cholesterol, total serum proteins, albumin and globulin.

puring necropsy examination of the external surface of the body, all orifices, and the cranial thoracic and abdominal cavities and their contents was performed. A thorough naked eye examination of size, shape, surface, colour, contours etc of all the important organs and tissues was done. Liver, kidneys, adrenals, brain heart lungs, spleen and gonads were registed, and their relative weights were also calculated.

All the above mentioned organs, and representative pieces from skin trachea, thyroid, different parts of the GIT pancreas, urinary bladder, mammary glands and whole eyes were preserved in 10% buffered formalin till they were processed for paraffin embedding and sectioning.

Representative places from all the preserved organs and tissues were processed for paraffin embedding. Four to six micrometer thick sections are cut and stained by haematoxylin and eosin according to standard methods for microscopic examination.

experimentation. Animals of both drug treated and control groups showed uniform (rats) or irregular (monkeys) trends of gain in body weight. The laboratory investigations showed some minor incidental variations but there was no indication of drug induced damages in the various urinalyses hematological and bloods biochemical values. Also, necropsy (including

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absolute and relative organ weights of important organs and histopathological examinations did not reveal any sign of target organ toxicity

Compound L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide was thus found safe in rats and rhesus monkeys in the 30-day toxicity study by subcutaneous route at the dose levels mentioned above.

EXAMPLE 7

Alleviation of pain:

Analgesia was measured by tail flick latencies (t.f.l.) assay in rats. (D'Amour, F. E and Smith., D. L., J. Pharmacol. Exp. Ther. 72,74-79, 1941) The test required determination of tail flick latencies to heat stimulus. The basal tail flick latencies were determined twice at 10 min interval and averaged to obtain single pre drug latency. An increase in the latency by 100% or more was indicative of analgesic state of animal. A cut-off time of 10 seconds was used to avoid damage to the tail skin.

groups of 8-10 rats each. Then tail flick latency was determined every 10 minutes till it was near the pre drug level. Percent of animals exhibiting analgesia was determined at each dose level of various compounds and the ED₅₀ along with 95% fiducial limits was calculated [Table 2] by prohibit analysis (Finney, 1971), The duration of analgesic effect was determined at the peat effect. The analgesia lasted for 5-6 hours after oral administration.

TABLE 2

	INDDE		
Compound/Formulation	ED ₅₀ (mg/Kg)		
Compound Formulation	S.C. route	P. O. route	
L-Tyrosyl-D-alanyl-glycyl- N-methylphenylalanyl-	2.58	22.30	
glycyl-isopropylamide L-Tyrosyl-D-alanyl-glycyl- N-methylphenylalanyl- glycyl-isopropylamide beta CD complex (1:2)	4.45 (equivalent to 0.89 mg of compound)	54.23 (equivalent to 10.84 mg of compound)	

It is evident from above table that the ED₅₀ of beta CD complex is about one third than the compound L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide by subcutaneous route and was one half by oral route. The LD₅₀ of the compound L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide is 1000 mg/Kg i.p.,and that of the complex is > 10,000 mg/kg P. O. in mice.

EXAMPLE 8

Anti-inflammatory action

Anti-inflammatory action was measured in vivo by the inhibition of oedema caused by carrageenin. Rats were given 250/500 mg/kg of the test substance 1 hour before the injection of 0.1 ml 1% carrageenin suspension. The inhibition of the formed oedema was measured 3 hours after injecting carrageenin. [Table 3].

TABLE 3

Measurement of anti-inflammatory action in vivo at the dosages of 500/250 mg/kg of the active substance applied per orally substance

	Dose (p.o.) (mg/kg)	Anti-inflammatory action (210 min)
L-Tyrosyl-D-alanyl-glycyl-N- methylphenylalanyl-glycyl-isopropyl amide:beta CD complex (1:2)	250	4.0
L-Tyrosyl-D-alanyl-glycyl-N- methylphenylalanyl-glycyl-isopropyl amide:beta CD complex (1:2)	500	18.4.
L-Tyrosyl-D-alanyl-glycyl-N- methylphenylalanyl-glycyl-isopropyl amide	50	19.6

The measurement in vivo of the anti-inflammatory action showed that the inclusion complex of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta cyclodextrin exhibited anti-inflammatory action.

EXAMPLE 9

Effect on gastric mucous membrane

Effect of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide and inclusion complex of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta cyclodextrin on gastric mucous membrane was measured in rats. Rats were fasted overnight and were given 100 mg/kg of the active substance orally. After 4 hours its effect on irritation of gastric mucous membrane and gastric ulceration, gastric juice volume and extent of free and total acid was determined. [Table 4].

Measurement of the effect of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide: beta-cyclodextrin (1:1) on gastric mucous membrane

isopropylamide:beta-cyclod	extrin (1:1) on gastric muc	2003 1110111072110	
		Free acid	Total acid
Dose (mg/kg) Ulceration	- 23	+ 9.0	+ 10.0
100			

The above data show that L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide bound into an inclusion complex with .beta -cyclodextrin did not exhibit an irritating effect on gastric mucous membrane of the animals.

EXAMPLE 10

Preparation of tablets with 125 mg of active substance [inclusion complex (1:2)] of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta-cyclodextrin

Tablets of the following composition were prepared:

TABLE 5

INDLE	
	Weight
Active constituent	125 mg
L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-	122 129
L-Tyrosyl-D-alalyl-gryoyl-1 thompolex (1:2) isopropylamide :beta cyclodextrin complex (1:2)	70
isopropylatilide .oca oyoloom	70 mg
lactose	5 mg
methyl cellulose	2 mg
magnesium stearate	

Preparation of tablets: The active substance was homogeneously stirred with additives. The mixture was sieved through a sieve and pressed into tablets on a rotating tableting machine and the resulting tablet properly packed in a polythene lined blister type packing to avoid coming into contact with moisture before use.

EXAMPLE 11

Preparation of transdermal tapes of active substance (inclusion complex (1:1)) of L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide with beta-cyclodextrin

Polyvinyl alcohol (4 g) and polyvinylpyrrolidone (2 g) were taken in a beaker and stirred 2 75° C, 120 ml ethyl alcohol (50% v/v) was slowly added into this to get a fine latex.

L-Tyrosyl-D-alanyl-glycyl-N-methylphenylalanyl-glycyl-isopropylamide beta- cyclodextrin complex (1:1) (1:8) g was taken & suspended in propylene glycol (2 ml) PEG 400 (0.5 ml),

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propylene glycol (2 ml) and tritan X-100 (1.5 ml) by using sonicator Polymer latex prepared in earlier step was transferred to it by continuous sterring and further stirred for 15 minutes. The hydrogel so formed was uniformly poured into a petri-dish and allowed to dry. The next day a yellow coloured patch was obtained. This was covered with the plastic film and cut into the desired size. The patches so obtained were properly packed in a polythene lined blister type packing to avoid coming into contact with maisture before use.